

An Alignment of 17 Deduced Protein Sequences from Plant, Fungi, and Ciliate H⁺-ATPase Genes

Achim Wach,¹ Alain Schlessler,¹ and André Goffeau¹

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Seventeen protein sequences of H⁺-ATPases from plants (*Arabidopsis thaliana*, *Nicotiana plumbaginifolia*, *Lycopersicon esculentum*), fungi (*Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Zygosaccharomyces rouxii*, *Neurospora crassa*, *Candida albicans*), and a parasitic ciliate (*Leishmania donovani*) have been aligned. Twenty sequence fragments were identified which were conserved in H⁺-, Na⁺/K⁺-, and Ca²⁺ plasma membrane-ATPases. In addition, a total of 118 residues not located in these fragments were found to be conserved in all H⁺-ATPases. Among those, 38 amino acid residues were screened out as being priority targets for site-directed mutagenesis experiments aimed at the identification of the amino acid residues specifically involved in cation specificity.

KEY WORDS: Yeast; proton ATPase; sequence alignment; hydrophobic regions.

INTRODUCTION

That the yeast plasma membrane H⁺-ATPase, initially identified by physiological studies (see review by Goffeau and Slayman, 1981), is structurally similar to the mammalian Ca²⁺- and Na⁺/K⁺-ATPases has been inferred from the observation of a predominant 100-kDa polypeptide in an ATPase solubilized and purified from the plasma membranes of the fission yeast *Schizosaccharomyces pombe* (Dufour and Goffeau, 1978). A similar subunit composition has been reported for the transport ATPase enzyme isolated from the mold *Neurospora crassa* (Bowman *et al.*, 1981; Addison and Scarborough, 1981) and from the budding yeast *Saccharomyces cerevisiae* (Malpartida and Serrano, 1980; Foury *et al.*, 1981; Koland and Hammes, 1986; Perlin *et al.*, 1989; Wach *et al.*, 1990). The biochemical characterization of the three fungal enzymes (see review by Goffeau and Slayman, 1981) has established the electrogenic H⁺-pumping capacity of the reconstituted purified enzyme and the similarity in

mechanisms with the mammalian Ca²⁺- and Na⁺/K⁺-ATPases. In particular, the presence of an aspartyl-phosphate catalytic intermediate and the sensitivity of the ATPase activity to vandate were established for the three fungal enzymes. A similar class of plant H⁺-ATPase was reported more recently (see review by Serrano, 1989).

Since 1984, at least 16 reviews reporting further properties of H⁺-ATPases from fungi and plant plasma membranes have been published (Serrano, 1984; Scarborough, 1985; Bowman and Bowman, 1986; Goffeau and Boutry, 1986; Goffeau, 1987; Slayman, 1987; Goffeau, 1988; Serrano, 1988; Goffeau *et al.*, 1989; Nakamoto and Slayman, 1989; Serrano, 1989; Goffeau and Green, 1990; Serrano, 1991; Gaber, 1992; Scarborough, 1992).

Among this wealth of largely redundant information, at least two alignments of fungal and plant H⁺-ATPase protein sequences have been previously proposed (Serrano, 1989; Goffeau and Green, 1990). However, during these last three years several new genes encoding presumed H⁺-ATPases from plant and fungi have been reported but not yet listed and compared to each other. Therefore in the present

¹Unité de Biochimie Physiologique, Université Catholique de Louvain, Place Croix du Sud 2-20, B-1348 Louvain-la-Neuve, Belgium.

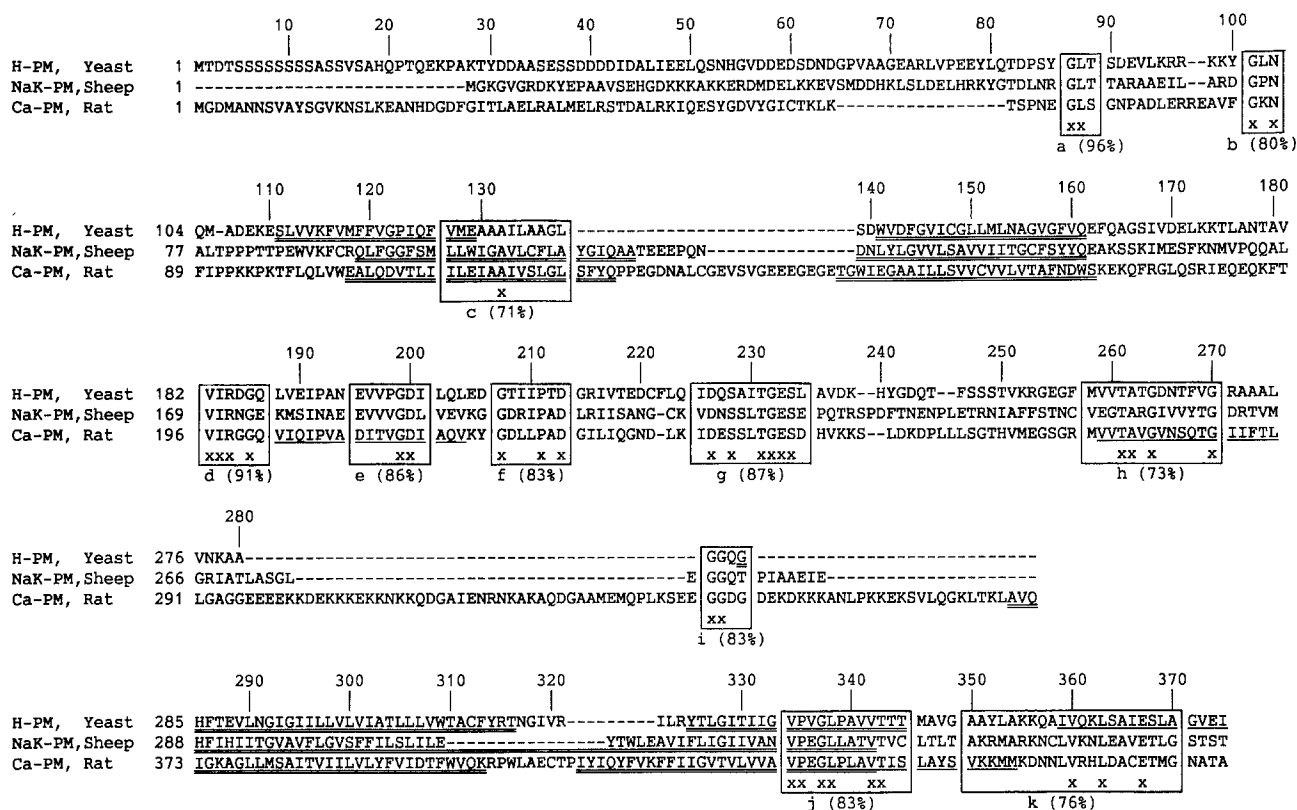


Fig. 1. Alignment of P-type ATPase sequences. Amino acid sequences are from *Saccharomyces cerevisiae* H⁺-ATPase (H-PM, Yeast), Na⁺/K⁺-ATPase from sheep (NaK-PM, Sheep), and Ca²⁺-ATPase from rat plasma membranes (Ca-PM, Rat). See text for references. The alignment was initially performed by using the program CLUSTAL (Higgins and Sharp, 1988) and then refined manually. The numbers on top of each block are positions in ScPMA1. Overall homologies were calculated by pairwise comparison of the sequences in each cluster using the structure-genetic matrix from Feng, *et al.* (1985). The clusters with mean homology scores above 70% are boxed and labeled a to s. Underlined sequences indicate predicted hydrophobic regions by the method of Klein *et al.* (1985); membrane integral sequences are underlined twice, and possibly membrane integral sequences are underlined once.

mini-review we will focus on the alignment of the 17 deduced H⁺-ATPase complete protein sequences reported, to our knowledge, up to January 1992. This will provide up-to-date information which might be useful for decisions concerning future site-directed mutagenesis or domain-shuffling experiments.

IDENTIFICATION OF 20 CONSERVED REGIONS IN THE Ca²⁺, Na⁺/K⁺, AND H⁺-ATPases

Green (1989) has identified about 30 amino acid sequence segments which share more than 15% mean identity in pairwise comparisons between skeletal Ca²⁺-ATPase versus sheep Na⁺/K⁺-ATPase, of sheep Na⁺/K⁺-ATPase versus yeast H⁺-ATPase, and of yeast H⁺-ATPase versus *E. coli* K⁺-ATPase (see also Goffeau and Green, 1990 and Van Dyck *et al.*, 1990).

Figure 1 represents an alignment, first performed by the CLUSTAL program (Higgins and Sharp, 1988) and subsequently refined manually, of three different eukaryotic plasma membrane ATPases pumping either Ca²⁺ (Schull and Greb, 1988), Na⁺/K⁺ (Schull *et al.*, 1985), or H⁺ (Serrano *et al.*, 1986, as corrected by Van Dyck *et al.*, 1990). The 20 sequence fragments showing a mean homology higher than 70% as calculated by pairwise comparison of the amino acid clusters quantified by the structure-genetic matrix from Feng *et al.* (1985) are boxed and marked by the letters a to s. The length of the homologous segments a to s varies from 3 to 39 amino residues, and the mean percent homology varies from 71 to 96%. Since these 20 segments a to s are well conserved in all three pumps, they are not likely to be involved in cation specificity but rather in the mechanism of ATP hydrolysis or in the mechanism of energy transduction which is considered to be basically similar for all three transport-ATPases.

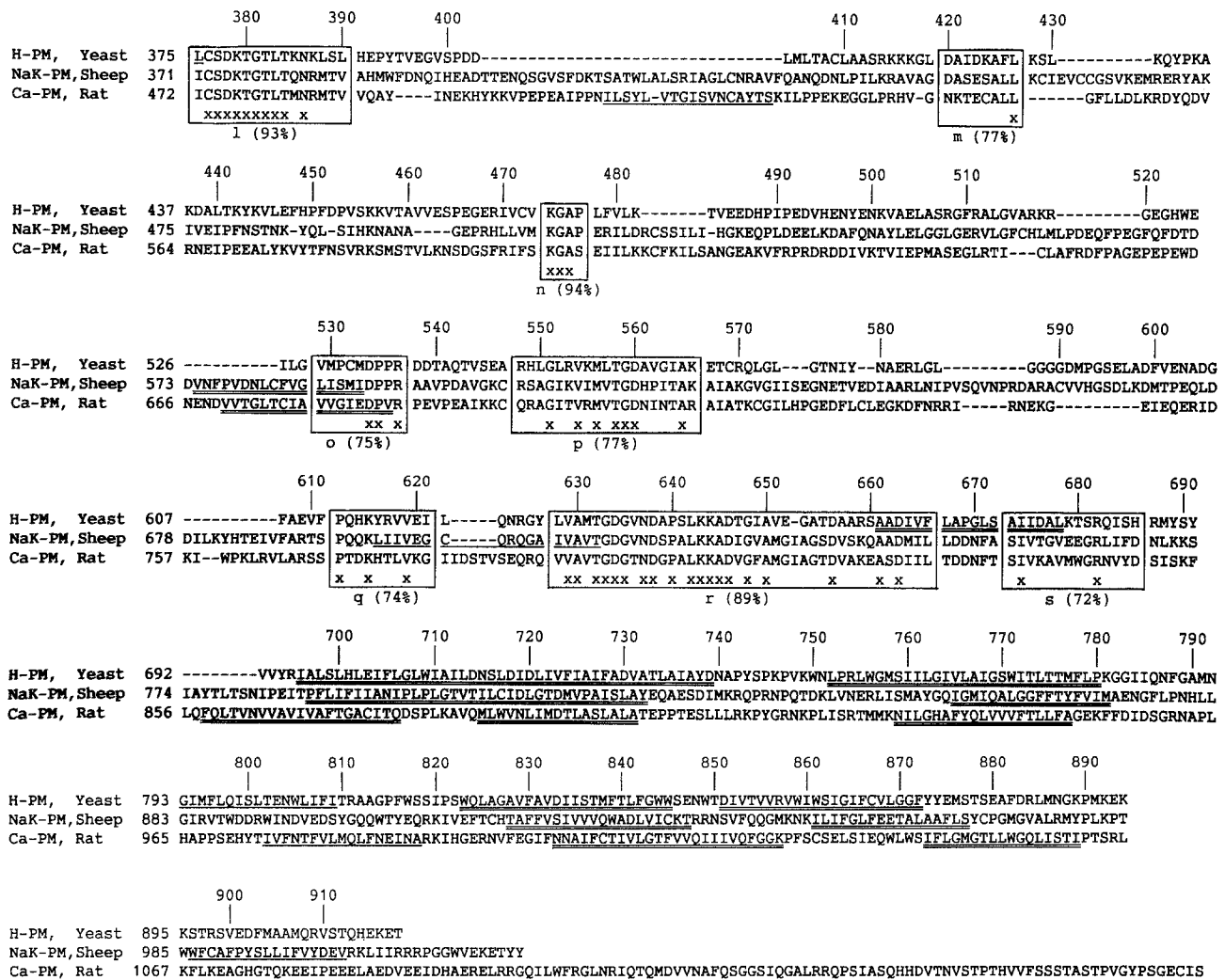


Fig. 1. Continued.

ALIGNMENT OF 17 AMINO ACID SEQUENCES FROM PUTATIVE H⁺-ATPases

Since the elucidation of the nucleotide sequence of the H⁺-ATPase gene from *Saccharomyces cerevisiae* (Serrano *et al.*, 1986; Van Dyck *et al.*, 1990; Perlin *et al.*, 1989) and *Neurospora crassa* (Addison, 1986; Hager *et al.*, 1986), numerous additional H⁺-ATPase gene sequences have been reported from fungi such as *Schizosaccharomyces pombe* (Ghislain *et al.*, 1987; Ghislain and Goffeau, 1991), *Saccharomyces cerevisiae* (Schlessler *et al.*, 1988), *Candida albicans* (Monk *et al.*, 1991), *Zygosaccharomyces rouxii* (Watanabe *et al.*, 1991), as well as from the protozoal ciliate *Leishmania donovani* (Meade *et al.*, 1987) and plants such as *Nicotiana plumbaginifolia* (Boutry *et al.*, 1989; Perez *et al.*, 1992; Morieau and Boutry, personal communication),

Arabidopsis thaliana (Harper *et al.*, 1989, 1990; Pardo and Serrano, 1989; Houlne and Boutry, personal communication), or *Lycopersicon esculentum* (Ewing *et al.*, 1990). To our knowledge, a total of at least 17 entire sequences of presumed H⁺-ATPase genes have been carried out from 1986 to 1991.

As seen in Fig. 2, the seven fungal ATPases align each other extremely well. The alignment of the nine plant ATPases is even more spectacular. If one excludes the first 31 to 172 (according to the species) N-terminal residues and the last 43 to 104 (according to the species) C-terminal residues, it is easy to align the plant and the fungal ATPase by introduction of 11 block deletions in the fungal sequences and 10 block deletions in the plant sequences. The deletions in fungal sequences ranging from 1 to 13 residues generally correspond to "insertions" in the plant sequences, and conversely the

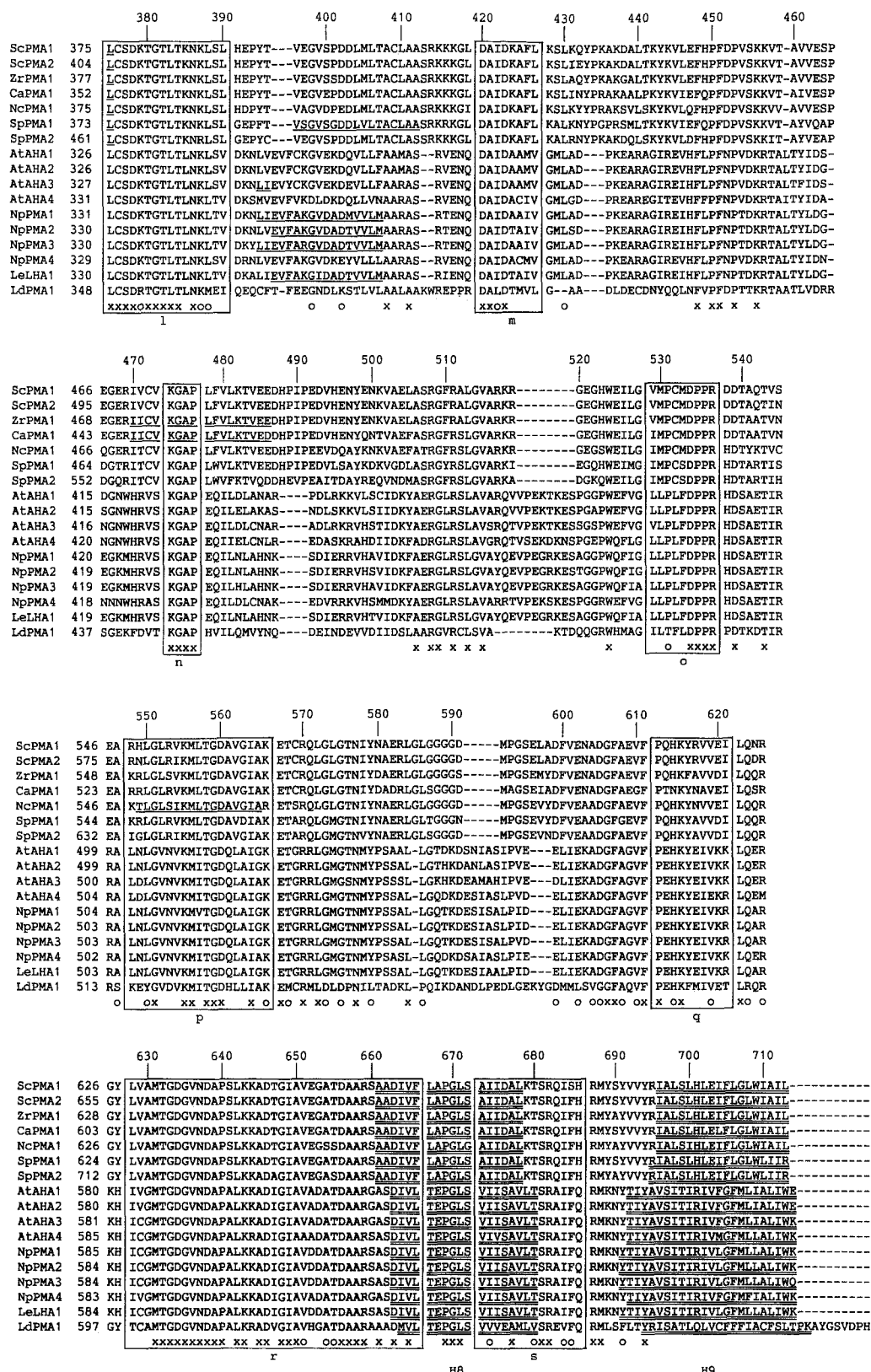


Fig. 2. Continued.

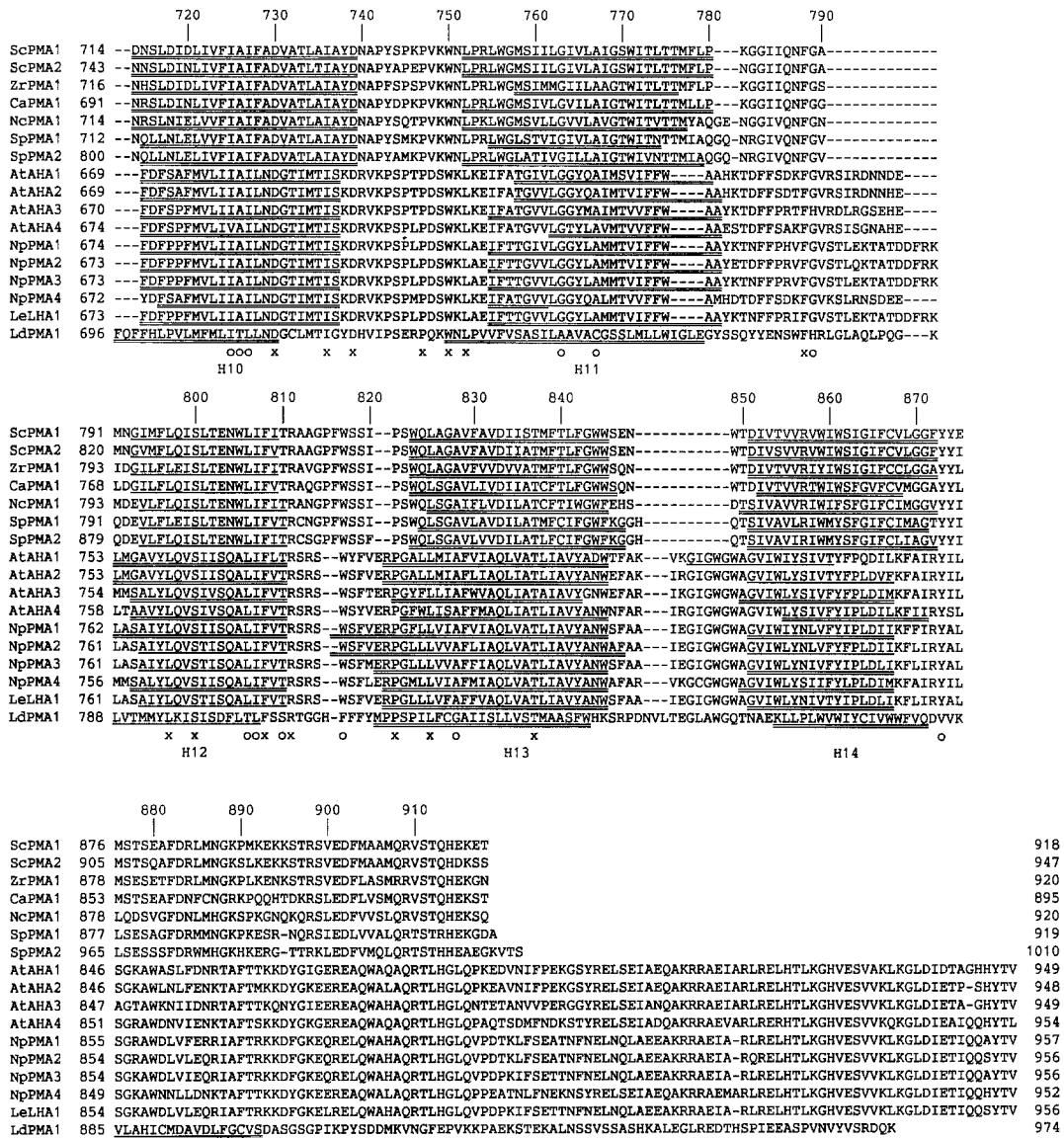


Fig. 2. Continued. Alignment of putative P-type H⁺-ATPase sequences. Amino acid sequences are from *Saccharomyces cerevisiae* (ScPMA1, ScPMA2), *Zygosaccharomyces rouxii* (ZrPMA1), *Candida albicans* (CaPMA1), *Neurospora crassa* (NcPMA1), *Schizosaccharomyces pombe* (SpPMA1, SpPMA2), *Arabidopsis thaliana* (AtAHA1-AtAHA4), *Nicotiana plumbaginifolia* (NpPMA1-NpPMA2), *Lycopersicon esculentum* (LeLHA1), *Leishmania donovani* (LdPMA1). See text for references. The alignment was initially performed by using the program CLUSTAL (Higgins and Sharp, (1988)) and then refined manually. The numbers on top of each block are positions in ScPMA1. Underlined sequences indicate predicted hydrophobic regions by the method of Klein *et al.* (1985): membrane integral sequences are underlined twice, and possibly membrane integral sequences are underlined once; these regions are labeled H1-H14. Conserved regions are derived from the sequence comparison in Fig. 1 are boxed and labeled from a to s. An x indicates total identity in all aligned sequences and a (o) indicates identity in all except one sequence. The overall identity is 13.3%, and the similarity is 24%.

plant “deletions” correspond to fungal insertions. These “insertions” and the N and C terminus represent a total of 22 segments which are not conserved among yeast and plants and which separate well-conserved domains. The evolutionary and functional

significances of these punctuations in the H⁺-ATPase genes have still to be clarified.

The *Leishmania donovani* sequence (Ld PMA1) is obviously distinct from the plant and the yeast H⁺-ATPase sequences. This raises the questions

whether or not it corresponds to a H⁺-ATPase as initially proposed by Meade *et al.* (1987) on the basis of sequence homologies.

A total of 241 residues identical in at least 16 out of 17 sequences (to allow for sequencing or alignment errors) is marked by an asterisk or a circle in Fig. 2. As expected, many of these are located in the boxed regions *a* to *s* which are conserved in ATPases of different cation specificity. However, 118 residues were found identical in at least 16 presumed H⁺-ATPases but not located in the conserved region *a* to *s*. This approach is prone to errors since the alignment depends partially on personal judgment. Nevertheless, it is remarkable that more than 100 identical residues can be identified which seem specifically conserved in the H⁺-ATPase sequences determined from different species: five fungi, three plants, and one ciliate.

PREDICTIONS OF HYDROPHOBIC REGIONS

The number and location of the membrane-spanning segments of the H⁺-ATPases are not known. Nevertheless, different topographies have been proposed for the yeast H⁺-ATPase comprising either 5, 8, 9, or 10 membrane spans (Hager *et al.*, 1986; Serrano *et al.*, 1986; Serrano, 1988, 1989; Davis and Hammes, 1989; Goffeau and Green, 1990). It is clearly established that the NH₂ and COOH termini are both located in the cytoplasm (Mandala and Slayman, 1989; Hennessey and Scarborough, 1990). This excludes the model of Serrano (1989) and Davis and Hammes (1989) who concluded on the existence of an uneven number of membrane spans. A biochemical investigation carried out recently (Subrahmanyeswara Rao *et al.*, 1991) concluded on the existence of up to 12 membrane spans. We are thus presently left with three possibilities: either 8, 10, or 12 transmembrane segments.

These discrepancies reflect the limitations of the different prediction methods for transmembrane segments. A recent study (Fasman and Gilbert, 1990) compared nine different predictive algorithms applied to the sequence of the reaction center from *Rhodospseudomonas viridis*, the molecular structure of which is known at 3 Å resolution (Deisenhofer *et al.*, 1985). It was concluded that the method with least ambiguity and highest accuracy in determining the membrane sequence is that of Klein *et al.* (1985). However, the method is not perfect: it misses the membrane span of

the H-polypeptide, and three amphiphilic regions of the L and M subunits with periphery/integral odds between 1.0 to 13.4 are predicted to be "possibly integral" but do in fact span the membrane as determined by X-ray crystallography.

We have applied the Klein *et al.* (1985) method to the detection of hydrophobic segments of the 17 putative H⁺-ATPase. This method determines 12–14 hydrophobic or amphiphilic segments in most sequences. For each gene product only 10 segments are generally predicted to be "integral" membrane segments, the others being predicted to be "possibly integral." These "integral" plus "possibly integral" segments are labelled H1 to H14 in Fig. 2. The segments H3 and H4 are considered "possibly integral" in plants and are predicted to be "peripheral" in yeasts. In contrast, H7 is predicted "possibly integral" in yeast and "peripheral" in plants. The segment H12 is predicted "possibly integral" in yeast and "integral" in plants. Only nine hydrophobic segments H1, H2, H5, H6, H8, H9, H10, H11, and H13 are predicted to be "integral" in all 17 sequences. The segment H14 might be put in the same category with 14 predictions as "integral" and three predictions as "possibly integral." These three later segments have peripheral/integral odds of 1.3, 4.0, and 21.6. Such odds are in the same order as those (from 2.5 to 13.4) given for the three amphiphilic membrane-spanning segments of the L and M subunits of the photosynthetic reaction center (Fasman and Gilbert, 1990). On this basis, we conclude on a consensus of 10 predicted membrane spans in all H⁺-ATPase as we had already proposed before (Goffeau and Green, 1990; Van Dyck *et al.*, 1990). It is, however, recognized that only straightforward biochemistry and structural data will be able to identify the real membrane topography.

SOME SUGGESTIONS FOR SITE-DIRECTED MUTAGENESIS

Site-directed mutagenesis of H⁺-ATPase turns out to be a more difficult approach than first believed. Even though ingenious procedures have been developed to put the wild type and/or mutant ATPase genes under the control of inducible promoters (Portillo and Serrano, 1988; Nakamoto *et al.*, 1991), it has not yet been possible to produce sufficient amounts of lethal mutant ATPase enzyme to carry out extensive biochemical studies. This is due to several reasons. The *S. cerevisiae* PMA1 gene is essential

Table I. Some Characteristic Residues Specifically Conserved in H⁺-ATPases from Yeasts and Plants^a

Domain	Boundaries (residue number in <i>S. cerevisiae</i>)	E or D	H	K or R	N or Q	P	W, F, or Y	S or T	C or M
Hydrophobic 1 (H ₁ and H ₂)	107 to 161	D107, <u>D143</u> , E162			N154	P123	F119		
Hydrophilic I	162 to 283	E190, E255		R215, <u>K252</u>				S249	
Hydrophobic 2 (H ₅ and H ₆)	284 to 345		H285				F286		
Hydrophilic II	346 to 660	D402, E567	H614		N577	P450, P453	Y579, F448 F607, F611	T543, T568	M688
Hydrophobic 3 (H ₈ to H ₁₄)	661 to 874	<u>D730</u> , D739		R687		P669, P747, P821	Y691, Y694, F788, W750	S800, T837	

^aThe residues underlined have been submitted to site-directed mutagenesis (see text and review by R. Gaber, 1992).

(Serrano *et al.*, 1986); therefore it is impossible to grow for more than a couple of divisions an ATPase mutant modified in an essential amino acid. Furthermore, a mixture of wild-type and lethal mutant ATPases is difficult to obtain in the same strain because of the presence of the PMA2 isogene which promotes intensive DNA repair of lethal mutations. Finally, no biochemical procedure exists today to separate purified wild-type from mutant ATPases. The very smart procedures developed by Nakamoto *et al.* (1991) for promoting specific accumulation of mutant enzyme into secretory vesicles might provide a solution to some of these problems but is probably not applicable to lethal mutation which might not be stably inserted in the endoplasmic reticulum. Anyway, an enormous number of possibilities of site-directed single amino acid substitutions exists: 918²⁰ for the H⁺-ATPase of *S. cerevisiae*. Choices have thus to be made. The alignment of 17 H⁺-ATPase of Fig. 2 might help in this respect. For instance, we suggest to give some priority to the residues listed in Table I. Our choice was based on the following rationale. One of the most interesting type of information we wish to obtain is the identification of residues specifically involved in H⁺-transport. Therefore we eliminated all residues contained in the segment *a* to *s* conserved in all eukaryotic transport-ATPases. We also did not consider the *Leishmania donovani* ATPase because of the uncertainty of it being a H⁺-ATPase and the numerous alignment ambiguities. Furthermore, we eliminated the N and C terminal as well as all the 20 "insertions" not conserved in both yeast and plant ATPases. We did not consider nonpolar residues such alanine, leucine, valine, isoleucine, and glycine because of their general unspecificity and unlikeliness to be

involved in H⁺-binding either by chelation or by direct protonation. Finally, among the remaining residues which were conserved in at least 15 of the plant and fungal ATPases, we eliminated those which were homologous to residues conserved in the Na⁺/K⁺- and Ca²⁺-ATPases in approximately similar position (with some flexibility to allow for misalignment) even when not located in the conserved region *a* to *s*. Using this rather empirical and partially subjective screen, we identified 38 residues: six residues in the hydrophobic region covering the predicted hydrophobic segments H1 and H2, five residues in the small hydrophilic domain, two residues in the predicted hydrophobic segments H5 and H6, 13 residues in the large hydrophilic domain, and 12 residues located in the hydrophobic region including the predicted hydrophobic segments H8-H14. Among the latter, the apparently specific conservation of three proline and four aromatic residues is rather surprising and not noticed so far to our knowledge. Among the 38 residues screened out by our approach, only three have been mutated so far (see review by Gaber, 1992). D143 and D730 were found to be essential, whereas K252 gives vanadate-resistant ATPase. It must be noted that the residue D730, which was proposed by Clarke *et al.* (1989) to be a H⁺-binding site similar to the Ca²⁺-binding site of the endoplasmic reticulum, was pinpointed by our method. This is only an example of the usefulness of our alignment. These alignments will become more precise and informative as additional sequences become available. New information analysis will have to be developed and will provide new structural and functional predictions. Unfortunately many of these cannot be tested yet biochemically, but it is to be hoped that a general method for expression

of lethal mutations of yeast H⁺-ATPase will become available in the near future.

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